

reproductive performance of male rabbits

**DOI:** <u>https://doi.org/10.36811/ojpsr.2021.110014</u> **OJPSR:** 

OJPSR: November-2021: Page No: 47-55

# Open Journal of Pharmaceutical Science and Research Review Article Open Access

Tramadol injection as dosage form -Induced deterioration in reproductive performance of male rabbits

Abtisam SA<sup>1</sup>, Osama HA<sup>2</sup>, Nosiba EB<sup>3</sup> and Fayrouz AK<sup>3\*</sup>

<sup>1</sup>Biomedical Science Department, Faculty of pharmacy, Omar Al-Mokhtar University, El -Beida-Libya <sup>2</sup>Biochemistry Department, Faculty of medicine, Omar Al-Mokhtar University, El -Beida-Libya <sup>3</sup>Chemistry Department, Faculty of Science, Omar Al-Mokhtar University, El -Beida-Libya

**\*Corresponding Author:** Fayrouz AK, Chemistry Department, Faculty of Science, Omar Al-Mokhtar University, El -Beida-Libya; Email: <u>fayalzobair@yahoo.com</u>

### Received Date: Oct 07, 2021 / Accepted Date: Nov 15, 2021 / Published Date: Nov 24, 2021

#### Abstract

Injections are usually more expensive than tablets, but they may be required less often since they are more concentrated, whereas a person may need to take a tablet more regularly, thus consuming a large volume. when we administer a drug via oral rout, its meant to be absorbed by stomach or intestinal mucosa, but its so happens that, gastric acid, food, other drugs and many other factor come into the play and retarded the drug absorption, metabolized in intestinal mucosal cells even before it can reach to liver or target tissue(first-pass metabolism)thus decrease the effective amount reaching to circulation, Now, a drug given through injection reaches circulation at cent percent level and show prompt action which is earnestly required at the moment, it also decrease the dose and systemic side effects . also the tablets formulation contain the excipient like filler, diluents for the purpose of long -term stabilization, bulking up tablet formulations making concentration of drug less in tablets and need more dose to give the therapeutic action. Tramadol is a potent analgesic effective in the treatment of mild to severe pains. However, the use of the drug can pose a threat to other organs and systems. Therefore, this study was designed to investigate the effect of tramadol injection as dosage form on reproductive parameters in male rabbits. Tenth rabbits were randomly divided into two equal groups (each group five rabbits). The first group was used as a control. The second group was used to study the effect of tramadol (50 mg/kg body weight) for six weeks. Results obtained showed that tramadol significantly (P<0.05) decreased libido (by increasing the reaction time), ejaculate volume, sperm concentration, total sperm output, sperm motility (%), total motile sperm per ejaculate (TMS), packed sperm volume (PSV), total functional sperm fraction (TFSF), normal and live sperm and semen initial fructose. While initial hydrogen ion concentration (pH) and dead and abnormal sperm were increased (P<0.05). Live body weight (LBW) and relative weights of testes (RTW) were significantly (P<0.05) decreased. Concentrations of thiobarbituric acid-reactive substances (TBARS) were significantly (P < 0.05) increased in plasma of rabbits treated with tramadol compared with control. The study showed the harmful effects of tramadol on the reproductive performance on male rabbits.

Keywords: Tramadol Injection; Rabbits; Semen; Testosterone



reproductive performance of male rabbits

DOI: https://doi.org/10.36811/ojpsr.2021.110014

OJPSR: November-2021: Page No: 47-55

**Cite this article as:** Abtisam SA, Osama HA, Nosiba EB, et al. 2021. Tramadol injection as dosage form -Induced deterioration in reproductive performance of male rabbits. Open J Pharm Sci Res. 3: 46-55.

**Copyright:** This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited. Copyright © 2021; Abtisam SA

### Introduction

Injections are commonly used when a quick result is required some medications may provide further benefits in addition to speed when injected. For example, a pain relief medication that's injected will better target the affected area, whereas a tablet will only provide general pain relief and may cause certain side effects, such as fatigue. A tablet pain reliever may also need to be taken regularly for a person to experience results, whereas the pain relief from an injection may last several weeks or months. Analgesics are a group of drugs that act in different ways on the central nervous system to analgesia and relief from pain [1]. In moderate and severe pain cases, opioid analgesic drugs are mainly used [2]. Opioid analgesic drugs might be obtained from natural sources (e.g. morphine, codeine) or from synthetic sources as Tramadol, heroin [3]. Tramadol hydrochloride is narcotic-like pain reliever drug centrally effective analgesic used for acute pain conditions include neuropathic, cancer and postoperative surgical pain [4].



Figure 1: Tramadol injection.

Nowadays, tramadol is widely used as an analgesic drug in human medicine [5]. Its mechanism of action is based on the inhibition of ascending pain to the central nervous system by its binding to µ- opiate receptors and inhibit the reuptake of norepinephrine and serotonin [6]. Continuous tramadol administration leads to the appearance of its toxic effects on various organs of the body [7]. Tramadol has toxic effects on the structure and function of hepatic, renal and testicular tissues of male albino rats [8]. In addition, the neurotoxicity of tramadol has been reported by [9]. High doses of tramadol cause neuronal degeneration in the rat brain [10] and alter brain neurotransmitter levels [11]. Tramadol can also bring on feelings of intense euphoria, which may be what prompts some individuals to start abusing it. Tramadol abuse among adolescents becomes a widely spread all over the world [12]. Generally, opioids are used as analgesic drugs without considering the several side effects already known. One of the side effects that is rarely considered is hypogonadism [13]. In recent times, it has been observed that intrathecal and oral opioids are capable of suppressing testosterone secretion throughout their period of administration [14]. Opioids, both endogenous and exogenous, modulate gonadal function primarily by acting on opioid receptors in the hypothalamus [15], inducing the decreased release or disruption of the normal pulsatility of gonadotropin releasing hormone secretion. This results in a reduction in the release of the luteinizing hormone (LH) and follicle-stimulating hormone (FSH) from the pituitary gland and of testosterone or estradiol (E2) from the gonads. Opioids can also have direct effects on the pituitary gland and the testes [16]. Tramadol abuse among



reproductive performance of male rabbits

DOI: https://doi.org/10.36811/ojpsr.2021.110014

OJPSR: November-2021: Page No: 47-55

adolescence has many health and social consequences Adolescence is a critical period for neurodevelopment. Exposure to addictive substances during this period leads to various alterations in brain histological structure that can be translated into functional consequences throughout life [17]. Although tramadol is a weak opioid, its long-term use may result in wide psychiatric and physical symptoms. The chronic administration of tramadol is associated oxidative inhibition with stress, of neurogenesis, apoptosis and mitochondrial dysfunction [18]. The frontal lobe control emotions, behavior, attention, judgment and plays a key role in future planning including self-management and decision-making Tramadol abuse lead to behavior and emotional disturbances, loss of confidence, neglect healthy social interactions and education regression [19]. The present study was designed to study the effects of tramadol in male rabbits on the reproductive performance.

## **Materials and Methods**

In this study tramadol was purchased from pharmacy alsalam hospital in El -Beida-Libya. The dosage of tramadol was 50 mg/kg BW each other day [20]. The animals were weighted and received the calculated dose of the drug according to their weight. Male New Zealand White rabbits (age of 6 months and initial weight of 1.892±50.79kg) were used. Animals were individually housed in cages and weighed weekly throughout 6-weeks experimental period. Rabbits nourished pellets which comprised of 30% berseem (Trifolium alexandrinum) roughage, 25% yellow corn, 26.2% wheat bran, 14% soybean supper, 3% molasses, 1% CaCl<sub>2</sub>, 0.4% NaCl, 0.3% blend of minerals and vitamins, and 0.1% methionine. The vitamin and mineral premix per kg contained the taking after IU/gm for vitamins or minerals: vit A-4000,000, vit D3-5000, 000, vit E-16,7 g, K-0.67 g, vit B1-0.67 g, vit B2-2 g, B6-0.67 g, B12-0.004 g, B5-16.7 g, Pantothinc acid-6.67 g, Biotein-0.07 g, Folic acid-1.67 g, Choline chloride-400 g, Zn-23.3 g,

Mn-10 g, Fe-25 g, Cu-1.67 g, I-0.25 g, Se-0.033g, and Mg-133.4 g (Rabbit premix created by Holland Nourish Connect. Co.).The chemical analysis of the pellets [19] showed that they contained 15.8 % crude protein, 11.3 % crude fiber, 3.7 % ether extract, 7.2 % ash, 92.9 % organic matter and 62.4 % nitrogen free extract % as DM basis.

A total of 10 mature male rabbits were randomly divided into two equal group. Group 1 served as control, while groups 2 was given tramadol (50 mg/kg body weight). The doses of tramadol was calculated according to the animal's body weight on the week before dosing. The proper doses of tramadol for each animal were placed into a syringe that was inserted orally with the help of plastic tube directly into the oesopharyngeal region. Semen collection was done weekly and continued throughout the 6-week experimental period. Ejaculates were collected using an artificial vagina and a teaser doe. The volume of each ejaculate was recorded (utilizing a graduated collection tube) after takeoff of the gel mass. A weak eosin solution [20] was used for evaluation of sperm concentration by the improved Neubauer haemocytometer slide (GmbH+Co., Brandstwiete 4, 2000 Hamburg Germany). 11. and Add up to sperm yield calculated by increasing semen ejaculate volume and semen concentration. Determination of initial fructose concentration in seminal plasma was determined immediately after semen collection according to [21]. Assessments of dead and normal spermatozoa were performed using an eosin-nigrosine blue staining mixture [22]. The percentages of motile sperm were estimated by visual examination under low-power magnification (10×) using light microscope. Add up to number of motile sperm was calculated by duplicating the rate of motile sperm and add up to sperm vield. Reaction time was chosen as the diminutive of subjecting a doe to the buck until the completion of erection; it was measured in seconds.



reproductive performance of male rabbits

DOI: https://doi.org/10.36811/ojpsr.2021.110014

OJPSR: November-2021: Page No: 47-55

Starting hydrogen particle concentration (pH) was determined immediately after collection using pH cooperative paper (Universalindikator pH 0-14 Merck, Merck KgaA, 64271 Darmstadt, Germany). Packed sperm volume (PSV) was recorded. Add up to useful sperm division (TFSF) was calculated as the item of add up to sperm yield, motility (%), and ordinary morphology (%) [23. Plasma thiobarbituric acid-reactive substances (TBARS) were measured by the strategy of [24. The relative weight of organs (%) was calculated as g/100 g body weight. Serum was obtained by centrifugation of blood samples at 860×g for 20 min, and was stored at  $(-20^{\circ C})$ until used for analysis Testosterone hormone assayed concentration were by using commercial kit that was supplied by Coat – A – Count testosterone RIA. from Diagnostic

Systems Laboratories (DSL), from Texas, USA. Statistical analysis: Where applicable, statistical analysis was carried out in Minitab software (version17) statistical significance was assessed using ANOVA analysis with Tukey multiple comparison test after detection normal distribution to the information and suitable P < 0.05 consider critical.

### Results

Observation of animal's tramadol-fed rabbits showed varying degrees of clinical signs few days after dose. The testicular weight in group tramadol was significantly lower than control group (Figure 1).



Figure 2: Morphology effect of tramadol after 6weeks on testes of male rabbits.

<b>Table 1:</b> The overall means (±SE) of be concentration and Thiobarbituric acid- treatment of male rabbits with tramadol	reactive substances (TI	
Parameters	Animal Groups	
	Control	Tramadol
BW (g)	$1892 \pm 50.79^{a}$	1529± 64.85°
RTW (g/100 g BW)	$4.432 \pm 0.486^{ab}$	$2.774 \pm 0.424^{b}$
Testosterone (ng/mL)	$1.570 \pm 0063^{b}$	$0987 \pm 0112^{\circ}$
TBARS (nmol/ml)	$2.673 \pm 0.025^{a}$	$3.113 \pm 0.087^{a}$
Testes	14.9±1.39 <sup>b</sup>	28.5±0.25ª
TBARS (nmol/g tissue)		
<sup>abc</sup> Within row, means with different sup	perscript letters differ sig	gnificantly (p<0.05).



reproductive performance of male rabbits

**DOI:** <u>https://doi.org/10.36811/ojpsr.2021.110014</u>

OJPSR: November-2021: Page No: 47-55

Body weight (BW) relative weight of testes and testosterone were significantly (P<0.05) decreased in rabbits treated with tramadol compared to control animals (Table 1). Results obtained showed that tramadol significantly (P<0.05) decreased libido (by increasing the reaction time), ejaculate volume, sperm concentration, total sperm output, sperm motility (%), total motile sperm per ejaculate (TMS), packed sperm volume (PSV), total functional sperm fraction (TFSF), normal and live sperm and semen initial fructose. While initial hydrogen ion concentration (pH) and dead and abnormal sperm were increased (P<0.05). Live body weight (LBW) and relative weights of testes (RTW) were significantly (P<0.05) decreased in plasma of rabbits treated with tramadol compared with control (Table 2).

**Table 2:** The overall means (±SE) of semen characteristics during treatment of male rabbits with tramadol.

Parameters	Animal Groups	
	Control	Tramadol
Ejaculate volume (ml)	$0.75 \pm 0.024^{ab}$	0.70±0.024 <sup>b</sup>
PH	7.84±0.032 <sup>b</sup>	8.08±0.074 <sup>a</sup>
Reaction time (s)	4.00±0.79 <sup>b</sup>	5.13±0.33 <sup>a</sup>
Packed sperm volume (%)	$14.6 \pm 0.17^{ab}$	13.34±0.22°
Sperm concentration ( $\times 10^6$ ml <sup>-1</sup> )	248±5.7 <sup>b</sup>	237±5.3 <sup>b</sup>
Total sperm output (×10 <sup>6</sup> )	185±6.5 <sup>b</sup>	166±6.9 <sup>b</sup>
Sperm motility (%)	66.2. ±0.95 <sup>b</sup>	62.5±1.1 <sup>b</sup>
Total motile sperm ( $\times 10^6$ )	123±5.0 <sup>b</sup>	104±4.6 <sup>b</sup>
Live sperm (%)	73.7±1.2 <sup>b</sup>	65.8±1.24 <sup>a</sup>
Dead sperm (%)	26.3±1.19 <sup>b</sup>	34.2±1.24 <sup>a</sup>
Normal sperm (%)	$82{\pm}0.5^{a}$	78±0.9 <sup>b</sup>
Abnormal (%)	18±0.2 <sup>b</sup>	22±06 <sup>a</sup>
Total functional sperm fraction (×10 <sup>6</sup> )	100±4.2 <sup>b</sup>	82±42°
Initial fructose (mg/dl)	264±5.2 <sup>b</sup>	229±5.8ª

#### Discussion

Tramadol addiction and abuse is now a major public health menace in libya. The rampancy of tramadol abuse and the consequential leap in crime in libya led to Government banning the manufacture, sale and use of tramadol. The design of the current study investigated the short- and long-term implications of tramadol abuse on semen quality indices using an animal model. Tramadol abuse has increased in the Middle East region, tramadol use was common among adolescents and over one third of tramadol users had drug-related problems [17]. This study has shown that the treatment of rabbits with tramadol caused significant reduction in body weight of rabbits at a later stage during the treatment period. This suggests that tramadol increases the catabolism of lipids in the adipose tissue, resulting in significant reduction in body weight of rabbits at a later stage during the treatment period. Similar results were reported by [25]in Persea americana leaf extractstreated rats. Results obtained revealed that tramadol treatments had significant effect on the body weight, sperm count and testicular integrity which agrees with the findings of [26-28] who reported that tramadol caused disorganization of the seminiferous tubules with almost missing of comparatively sperm and decreased spermatogenic cells. Results obtained are also in line with previous report on the gonadotoxic



reproductive performance of male rabbits

DOI: https://doi.org/10.36811/ojpsr.2021.110014

OJPSR: November-2021: Page No: 47-55

effects of tramadol in male animal models by [29.30] who reported that tramadol significantly decreased sex hormones and degeneration of spermatogonia, distortion of Sertoli cell tight junctions, and accumulation of electron-dense bodies in Sertoli cells. Histological findings clearly revealed that the normal architecture and integrity of the testes of tramadol treated animals were altered causing atrophy and necrosis of the spermatogenic cells (Sertoli and Leydig cells). Sertoli cells are considered as nursery units for the developing sperms [31]. Also, [32] reported that Sertoli cells are major supporters of spermatogenesis and germ cells because of the secretion of proteins such as core protein histone, androgen binding protein, and androgen binding proteinheat shock protein, Ncadherin, and desmoglein. The Levdig cells play an important role in the function and structure of seminiferous tubules and in the synthesis of testosterone, which is vital for the regulation of spermatogenesis. Reduced intra-testicular testosterone results in apoptosis of germ cells [33,34] added that steroid biosynthesis is a multistep process controlled by pituitary hormones, and this process is accelerated by the hormone dependent organelle communication network mediated by protein-to-protein interactions and inter-organelle trafficking, resulting in the efficient and timed delivery of cholesterol into the mitochondria for steroid synthesis. Therefore, reduced steroidogenesis results in altered spermatogenesis and spermatic failure. This could be the underlying cause of the significant reduction in the sperm count and weight of testes observed in tramadol treated animals. This assertion is corroborated by [35,36]. More so, [37] reported that the distortion in fertility in male mammals is directly correlated with the disruption of spermatogenesis and the hormone regulatory mechanisms and pathways.Also, Tramadol caused significant reduction in sperm motility. This suggest that the drug has an inhibitory effect on fertilizing capacity, since it has been reported that sperm motility is of importance with regard to sperm fertilizing capacity [38].

Similar report was given by 18 in rats treated with Pueraria tuberosa root extract. Youths are known to continue using these drugs despite the major risk behaviours with its accompanied physical and mental health complications [39].

Studies have also shown that increased reactive oxygen species (ROS) levels and oxidative stress correlates positively with decreased sperm parameters [40-42], where it reported that oxidative stress causes significant damage biological molecules such as lipid to peroxidation, DNA damage and testicular histopathology as well as decline in sperm quality. It has also been reported that tramadol suppresses testosterone by inducing nitric oxide (NO) [43]. A wellcharacterized consequence of compounds is the reduction NO in steroidogenic enzyme activities [44]. Inhibition of LH stimulated steroidogenesis may be reinforced by NO in Leydig cells [45]. Excessive NO production might inhibit the production of testicular adenosine 3, 5-cyclic monophosphate, which helps to transport the inner mitochondrial cholesterol to membrane, culminating in lower testosterone release [46]. All these reports suggest that the degenerative changes in germ cells observed in this study might be attributed to hormonal deficiency. significant decrease А in testosterone levels were seen in this study which correlates with [47] suggesting that long term opioid therapy results in clinically relevant suppression of both hypothalamopituitaryadrenal and –gonadal axes with suppression in testosterone, estrogen and cortisol. This effect as seen in this sub-acute study may be attributed to suppression of the hypothalamic-pituitarygonadal axis (HPG axis) by opioids [48,49]. Testosterone and estrogen (the male and female principal sex hormones) are produced by the testes and ovaries respectively by the action of FSH and LH (referred to as gonadotropins) which are produced by the pituitary gland following the stimulation of gonadotropinreleasing hormone (GnRH) which in turn is produced by the hypothalamus. It is expected that as the level of sex hormones rises



reproductive performance of male rabbits

DOI: <u>https://doi.org/10.36811/ojpsr.2021.110014</u> OJPSR: November-2021: Page No: 47-55

a negative feedback loop should trigger the hypothalamus to reduce production of GnRH.

### Conclusion

Parenteral products are liquid preparations administered directly into body tissues rather than via the oral route. "Parenteral" is itself derived from the Greek words para (beside) and enteron (the intestine) and most often refers to either subcutaneous (SC), intramuscular (IM), or intravenous (IV) administration of drugs. Liquid preparations administered by injection are characterised by three qualities possessed by no other type of pharmaceutical dosage form: sterility, freedom from pyrogens, and particulate matt. The findings of the study provide substantial evidence that tramadol injection as dosage form has an adverse effect on sperm profile and reproductive organs of male albino rabbits in terms of body weight, weight of testes, TBARS, sperm count, sperm viability, sperm motility as well as sperm head abnormalities, it is recommended that moderation should be exercised in the consumption of this drug by those taking it for therapeutic purpose.

### References

[1]. Kumar M, Shete A, Akbar Z. 2010. A review on analgesic: from natural sources. Int J Pharm Biol Arch. 1: 95-100.

[2]. El Shal EB, Selim MMH. 2015. The effect of tramadol treatment on rat testes and the possible protective role of selenium (light and electron microscopic study. Assiut Med J. 13: 127-137.

[3]. Hussein SA, Ismail HK, Abdel Aal SA. 2017. Effect of tramadol drug on some biochemical and immunological parameters in albino male rats; evaluation of possible reversal following its withdrawal. Benha veterinary medical journal. 33: 418-429.

[4]. Nagakannan P, Shivasharan BD, Thippeswamy BS. 2012. Effect of tramadol on behavioral alterations and lipid peroxidation after transient forebrain ischemia in rats. Toxicology mechanisms and methods. 22: 674-678. Ref.:

https://pubmed.ncbi.nlm.nih.gov/22871232/ DOI:

https://doi.org/10.3109/15376516.2012.716092

[5]. Azari O, Emadi L, Kheirandish R. 2014. The effects of long-term administration of tramadol on epididymal sperm quality and testicular tissue in mice. Iranian Journal of Veterinary Surgery. 9: 23-30.

[6]. Aldalou AR, Abdel AI, Shahwan O. 2014. Impact of giving sildenafil (viagra)/tramadol (tramal) combination on the blood of domestic rabbits. Journal of Science. 4: 162-169.

[7]. Shadnia S, Soltaninejad K, Heydari K. 2008. Tramadol intoxication: a review of 114 cases. Human & experimental toxicology. 27: 201-205. Ref.:

https://pubmed.ncbi.nlm.nih.gov/18650251/ DOI:

https://doi.org/10.1177/0960327108090270

[8]. Sayed HYM, Zidan AH. 2016. Histopathological and biochemical effects of acute and chronic tramadol drug toxicity on liver, kidney, and testicular function in adult male albino rats. J Forensic. 1: 41.

[9]. Hussein SA, Ismail HK, Abdel ASA. 2017. Effect of tramadol drug on some biochemical and immunological parameters in albino male rats; evaluation of possible reversal following its withdrawal. Benha veterinary medical journal. 33: 418-429.

[10]. Atici S, Cinel I, Cinel L. 2005. Liver and kidney toxicity in chronic use of opioids: an experimental long term treatment model. Journal of biosciences. 30: 245-252. Ref.:

https://pubmed.ncbi.nlm.nih.gov/15886461/ DOI: https://doi.org/10.1007/bf02703705

[11]. Bloms FP, Dremencov E, Cremers TIFH. 2011. Tramadol increases extracellular levels of serotonin and noradrenaline as measured by in vivo microdialysis in the ventral hippocampus of freely-moving rats. Neuroscience letters. 490: 191-195. Ref.:

https://pubmed.ncbi.nlm.nih.gov/21195741/ DOI:

https://doi.org/10.1016/j.neulet.2010.12.049



reproductive performance of male rabbits

DOI: https://doi.org/10.36811/ojpsr.2021.110014

OJPSR: November-2021: Page No: 47-55

[12]. Radbruch L, Glaeske G, Grond S. 2013. Topical review on the abuse and misuse potential of tramadol and tilidine in Germany. Substance abuse. 34: 313-320. Ref.: https://pubmed.ncbi.nlm.nih.gov/23844964/ DOI:

https://doi.org/10.1080/08897077.2012.735216

[13]. Reddy RG, Aung T, Karavitaki N. 2010. Opioid induced hypogonadism. BMJ. 341. Ref.:

https://pubmed.ncbi.nlm.nih.gov/20807731/ DOI: https://doi.org/10.1136/bmj.c4462

[14]. Aloisi AM, Ceccarelli I, Carlucci M. 2011. Hormone replacement therapy in morphine-induced hypogonadic male chronic pain patients. Reproductive Biology and Endocrinology. 9: 1-10. Ref.: https://pubmed.ncbi.nlm.nih.gov/21332999/

DOI: https://doi.org/10.1186/1477-7827-9-26

[15]. Vuong C, Van USH, O'Dell LE. 2010. The effects of opioids and opioid analogs on animal and endocrine systems. Endocrine reviews. 31: 98-132. Ref.:

https://pubmed.ncbi.nlm.nih.gov/19903933/ DOI: https://doi.org/10.1210/er.2009-0009

[16]. Adams ML, Sewing BRYAN, Forman JB. 1993. Opioid-induced suppression of rat testicular function. Journal of Pharmacology and Experimental Therapeutics. 266: 323-328. Ref.:

https://pubmed.ncbi.nlm.nih.gov/8392556/

[17]. Bassiony MM, Abdelghani M, Salah ED. 2018. Opioid use disorders attributed to tramadol among Egyptian university students. Journal of addiction medicine. 12: 150-155. Ref.:

https://pubmed.ncbi.nlm.nih.gov/29334513/ DOI:

https://doi.org/10.1097/adm.000000000000380

[18]. Barbera N, Fisichella M, Bosco A. 2013. A suicidal poisoning due to tramadol. A metabolic approach to death investigation. Journal of forensic and legal medicine. 20: 555-558. Ref.: https://pubmed.ncbi.nlm.nih.gov/23756535/ DOI:

### https://doi.org/10.1016/j.jflm.2013.03.006

[19]. El Wasify M, Fawzy M, Barakat D. 2018. The Sociodemographic and clinical characteristics of tramadol dependence among Egyptians and their relationship to the associated insomnia. Addictive Disorders & Their Treatment. 17: 98-106.

[20]. Ahmad RM, AL-Hubaity AY, Alazow NS. 2019. The Role of Vitamin C on the Structural Changes of Male Albino Rats Kidney Induced by Tramadol. Annals of the College of Medicine. 41: 57-62.

[21]. Mann T. 1948. Fructose content and fructolysis in semen. Practical application in the evaluation of semen quality. The Journal of Agricultural Science. 38: 323-331.

[22]. Blom E. 1950. A one-mintue live-dead sperm stain by means of eosin-nigrosin. J Fertil Steril. 1: 176-177.

[23]. Correa JR, Zavos PM. 1996. Preparation and recovery of frozen-thawed bovine spermatozoa via various sperm selection techniques employed in assisted reproductive technologies. Theriogenology. 46: 1225-1232. Ref.:

https://pubmed.ncbi.nlm.nih.gov/16727985/

DOI: <u>https://doi.org/10.1016/s0093-</u> 691x(96)00293-2

[24]. Tappel AL, Zalkin H. 1959. Lipide peroxidation in isolated mitochondria. Archives of Biochemistry and Biophysics. 80: 326-332.

[25]. Brai BIC, Odetola AA, Agomo PU. 2007. Effects of Persea americana leaf extracts on body weight and liver lipids in rats fed hyperlipidaemic diet. African journal of Biotechnology. 6.

[26]. El Sawy MM, Malak HWA. 2015. Effect of tramadol abuse on testicular tissue of adult albino rats: a light and electron microscopic study. Egyptian Journal of Histology. 38: 356-366.

[27]. Ceccarelli I, Rossi A, Maddalena M. 2009. Effects of morphine on testosterone levels in rat C6 glioma cells: modulation by anastrozole. Journal of cellular physiology. 221: 1-4. Ref.:

https://pubmed.ncbi.nlm.nih.gov/19492405/ DOI: https://doi.org/10.1002/jcp.21830



reproductive performance of male rabbits

**DOI:** <u>https://doi.org/10.36811/ojpsr.2021.110014</u> **OJPSR:** 

OJPSR: November-2021: Page No: 47-55

[28]. El Ghawet HA. 2015. Effects of tramadol on the reproductive function of wistar albino rats. Eur J Exp Biol. 5: 56-64.

[29]. Ekaluo UB, Udokpoh AE, Ikpeme EV. 2008. Effect of chloroquine treatments on sperm count and weight of testes in male rats. Global Journal of Pure and Applied Sciences. 14: 175-177.

[30]. El-Gaafarawi II. 2006. Biochemical toxicity induced by tramadol administration in male rats. The Egyptian Journal of Hospital Medicine. 23: 353-362.

[31]. Sawada H, Esaki M. 2003. Electron microscopic observation of 137 Cs-irradiated rat testis: production of basal laminae for germ cells, despite their absence. Microscopy. 52: 391-397. Ref.:

https://pubmed.ncbi.nlm.nih.gov/14599101/ DOI: https://doi.org/10.1093/jmicro/52.4.391

[32]. Mruk DD, Cheng CY. 2004. Sertoli-Sertoli and Sertoli-germ cell interactions and their significance in germ cell movement in the seminiferous epithelium during spermatogenesis. Endocrine reviews. 25: 747-806. Ref.:

https://pubmed.ncbi.nlm.nih.gov/15466940/ DOI: https://doi.org/10.1210/er.2003-0022

[33]. Ge R, Chen G, Hardy MP. 2009. The role of the Leydig cell in spermatogenic function. Molecular Mechanisms in Spermatogenesis. 255-269. Ref.: <u>https://pubmed.ncbi.nlm.nih.gov/19856172/</u> DOI: <u>https://doi.org/10.1007/978-0-387-</u> 09597-4 14

[34]. Issop L, Rone MB, Papadopoulos V. 2013. Organelle plasticity and interactions in cholesterol transport and steroid biosynthesis. Molecular and cellular endocrinology. 371: 34-46. Ref.: https://pubmed.ncbi.nlm.nih.gov/23246788/ DOI:

https://doi.org/10.1016/j.mce.2012.12.003

[35]. Ax RL, Collier RJ, Lodge JR. 1976. Effects of dietary caffeine on the testis of the domestic fowl, Gallus domesticus. Reproduction. 47: 235-238. Ref.: https://pubmed.ncbi.nlm.nih.gov/957321/ DOI: https://doi.org/10.1530/jrf.0.0470235 [36]. Ezzat AR, El-Gohary ZM. 1994. Hormonal and histological effects of chronic caffeine administration on the pituitary-gonadal and pituitary-adrenocortical axes in male rabbits. Functional and developmental morphology. 4: 45-50. Ref.:

https://pubmed.ncbi.nlm.nih.gov/7819609/

[37]. Glover A, Assinder SJ. 2006. Acute exposure of adult male rats to dietary phytoestrogens reduces fecundity and alters epididymal steroid hormone receptor expression. Journal of endocrinology. 189: 565-573. Ref.:

https://pubmed.ncbi.nlm.nih.gov/16731787/ DOI: https://doi.org/10.1677/joe.1.06709

[38]. Aitken RJ. 1984. A prospective study of the relationship between semen quality and fertility in cases of unexplained infertility. Journal of andrology. 5: 297-303. Ref.:

https://pubmed.ncbi.nlm.nih.gov/6540770/

DOI: <u>https://doi.org/10.1002/j.1939-</u> 4640.1984.tb00792.x

[39]. Mamman H, Othman AT, Lian LH. 2014. Adolescent's and drugs abuse in Nigeria. Journal of biology, agriculture and healthcare. 4: 5-9.

JS, [40]. Armstrong Rajasekaran M. Chamulitrat W. 1999. Characterization of reactive oxygen species induced effects on human spermatozoa movement and energy metabolism. Free Radical Biology and Medicine. 26: 869-880. Ref.: https://pubmed.ncbi.nlm.nih.gov/10232830/

DOI: <u>https://doi.org/10.1016/s0891-</u> 5849(98)00275-5

[41]. Agarwal A, Prabakaran SA. 2005. Mechanism, measurement, and prevention of oxidative stress in male reproductive physiology. 43: 963-974.Ref.: https://pubmed.ncbi.nlm.nih.gov/16315393/

[42]. Makker K, Agarwal A, Sharma R. 2009. Oxidative stress & male infertility. Indian Journal of Medical Research. 129: 357. Ref.: https://pubmed.ncbi.nlm.nih.gov/19535829/

[43]. Ahmed MA, Kurkar A. 2014. Effects of opioid (tramadol) treatment on testicular functions in adult male rats: The role of nitric



reproductive performance of male rabbits

DOI: <u>https://doi.org/10.36811/ojpsr.2021.110014</u> OJPSR: Nove

OJPSR: November-2021: Page No: 47-55

oxide and oxidative stress. Clinical and Experimental Pharmacology and Physiology. 41: 317-323. Ref.: https://pubmed.ncbi.nlm.nih.gov/24472030/ DOI: https://doi.org/10.1111/1440-1681.12213 [44]. Kostic TS, Andric SA, Maric D. 2000. Inhibitory effects of stress-activated nitric oxide on antioxidant enzymes and testicular steroidogenesis. The Journal of steroid biochemistry and molecular biology. 75: 299-306. Ref.:

https://pubmed.ncbi.nlm.nih.gov/11282286/ DOI: https://doi.org/10.1016/s0960-0760(00)00185-0

[45]. Dobashi M, Fujisawa M, Yamazaki T. 2001. Inhibition of steroidogenesis in Leydig cells by exogenous nitric oxide occurs independently of steroidogenic acute regulatory protein (star) mRNA. Archives of andrology. 47: 203-209. Ref.:

https://pubmed.ncbi.nlm.nih.gov/11695844/ DOI:

https://doi.org/10.1080/014850101753145915

[46]. Guo CH, Huang CJ, Chen ST. 2001. Serum and testicular testosterone and nitric oxide products in aluminum-treated mice. Environmental toxicology and pharmacology. 10: 53-60. Ref.: https://pubmed.ncbi.nlm.nih.gov/11382556/

[47]. Ballantyne JC. 2007. Opioid analgesia: perspectives on right use and utility. Pain physician. 10: 479. Ref.: https://pubmed.ncbi.nlm.nih.gov/17525783/

[48]. Aloisi AM, Ceccarelli I, Fiorenzani P. 2010. Aromatase and 5-alpha reductase gene expression: modulation by pain and morphine treatment in male rats. Molecular Pain. 6: 1744-8069. Ref.:

https://pubmed.ncbi.nlm.nih.gov/20977699/ DOI: https://doi.org/10.1186/1744-8069-6-69

[49]. Colameco S, Coren JS. 2009. Opioidinduced endocrinopathy. Journal of Osteopathic Medicine. 109: 20-25. Ref.: https://pubmed.ncbi.nlm.nih.gov/19193821/